

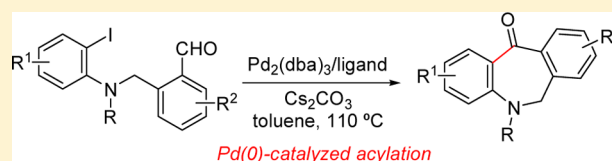
Synthesis of Dibenzo[*b,e*]azepin-11-ones by Intramolecular Palladium-Catalyzed Acylation of Aryliodides with Aldehydes

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S Supporting Information

ABSTRACT: The paper describes an efficient methodology for the synthesis of diversely functionalized dihydrodibenzo[*b,e*]azepin-11-ones based on the Pd(0)-catalyzed intramolecular acylation of aryl iodides with aldehydes.

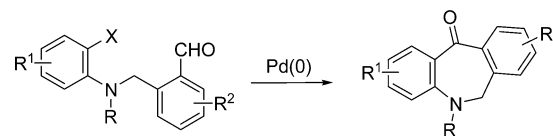


Palladium-catalyzed coupling reactions are well-established as reliable carbon–carbon bond-forming tools both in the research laboratory and in large-scale industrial production.¹ Among the variety of available cross-coupling processes, the palladium-catalyzed nucleophilic additions of aryl halides^{2–8} and arylboronic acids^{9–13} to carbon–heteroatom multiple bonds have been attracting increasing interest in recent years and are now emerging as powerful synthetic methodologies.

As part of our ongoing research on the synthesis of nitrogen heterocycles by means of palladium-catalyzed intramolecular coupling reactions of aryl halides and carbonyl compounds,¹⁴ we have been studying the possibility of controlling the ambiphilic character of the σ -arylpalladium species involved in these reactions.^{7,8,15,16} In this context, we have recently reported that when starting from (2-iodoanilino) aldehydes, both the enolate arylation and the acylation of the aryl halide can also be selectively promoted by Pd(0) with only a slight modification of the reaction conditions.¹⁷ The latter acylation reaction seems to proceed through a mechanism involving a carbopalladation between the σ -arylpalladium(II) moiety and the carbonyl group, followed by a β -hydride elimination from the resulting Pd(II) alkoxide intermediate. Besides its mechanistic implications, this reaction has considerable synthetic interest because it could constitute an alternative to classical acylation methodologies such as the Friedel–Crafts reaction, whose intrinsic problems of regioselectivity often decrease its effectiveness in the synthesis of unsymmetrically substituted derivatives.¹⁸ Moreover, it also provides an alternative to our previously reported palladium-catalyzed acylation of aryl halides by esters⁷ and amides.⁸ As these palladium-catalyzed reactions involve the nucleophilic attack of the σ -arylpalladium(II) species at the carbonyl group, the use of the more electrophilic aldehydes as the acylating agents would result in a more efficient methodology.

To demonstrate the potential of the palladium-catalyzed acylation reaction with aldehydes,^{3,19} we envisioned the synthesis of polycyclic compounds containing the dibenzo[*b,e*]azepine moiety (Scheme 1). The dibenzo[*b,e*]azepine system has attracted considerable attention due to its

Scheme 1



ubiquitous presence in the molecular structure of important bioactive compounds.²⁰ Therefore, the development of reliable methodologies for the construction of the dibenzo[*b,e*]azepine skeleton would be highly useful in the field of synthetic organic chemistry.²¹ In particular, palladium-catalyzed annulation reactions have provided some attractive and valuable routes for the synthesis of dibenzo[*b,e*]azepine derivatives.²²

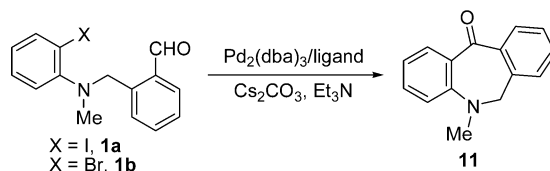
We commenced our investigation by studying the acylation reactions of aldehydes **1a** and **1b** (Table 1). We were pleased to find that starting from iodoaniline **1a**, the acylation can be smoothly promoted by using 5 mol % of $Pd_2(dba)_3$, 10 mol % of the phosphine ligand, and Cs_2CO_3 as the base in toluene at high temperature. As also observed in our initial studies with (2-iodoanilino) aldehydes,¹⁷ sterically hindered phosphines (i.e., dtpf, tBu_3P , and (*o*-tolyl)₃P) proved to be highly efficient ligands for the acylation and afforded dihydrodibenzo[*b,e*]azepin-11-one **11**²³ in a 75% average yield (Table 1, entries 1–3). The use of the same combination of catalyst and base in either acetonitrile or DMF was found to be less effective in promoting the acylation reaction (entries 4 and 5). Bromide **1b** was less efficient than iodide **1a** in the annulation reaction, and longer reaction times were required to reach the total conversion of the starting material (entries 6–8).

Although the acylation can be carried out effectively in toluene under simple thermal heating, the long reaction times required for the full conversion of the starting material, especially when starting from bromide **1b**, prompted us to also explore the reaction under microwave irradiation (entries 9–13).

Received: May 3, 2013

Published: July 30, 2013

Table 1. Optimization of Acylation Reaction Conditions



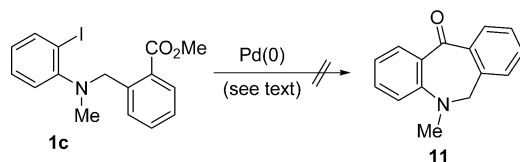
entry	aldehyde	ligand	solvent	time	¹ H NMR ratio	yield (%) ^a
1	1a	dtpf	toluene ^b	24 h		11 (77%)
2	1a	(^t Bu) ₃ P·HBF ₄	toluene ^b	24 h		11 (71%)
3	1a	(<i>o</i> -tolyl) ₃ P	toluene ^b	36 h		11 (75%)
4	1a	dtpf	CH ₃ CN ^b	24 h		11 (32%)
5	1a	dtpf	DMF ^b	24 h		11 (34%)
6	1b	dtpf ^c	toluene ^b	36 h	1b:11 (5:1)	
7	1b	(^t Bu) ₃ P·HBF ₄ ^c	toluene ^b	72 h	1b:11 (1:1.1)	
8	1b	(<i>o</i> -tolyl) ₃ P ^c	toluene ^b	120 h		11 (48%)
9	1a	dtpf	toluene ^d	60 min	1a:11 (3:1)	
10	1a	dtpf	dioxane ^d	60 min	1a:11 (3.5:1)	
11	1a	dtpf	DMF ^d	30 min		11 (35%)
12	1b	dtpf	DMF ^d	30 min		11 (37%)
13	1b	dtpf	CH ₃ CN ^d	30 min		11 (34%)

^aYields refer to pure products isolated by flash chromatography. ^bThe reactions were carried out using Pd₂(dba)₃ (0.05 equiv), ligand (0.1 equiv), Cs₂CO₃ (3 equiv), and Et₃N (6 equiv) in the indicated solvent at 110 °C in a sealed tube. ^c0.075 equiv of Pd₂(dba)₃ and 0.15 equiv of ligand were used. ^dThe reactions were carried out using Pd₂(dba)₃ (0.05 equiv), ligand (0.1 equiv), Cs₂CO₃ (1 equiv), and Et₃N (3 equiv) in the indicated solvent at 120 °C in a sealed vessel in a microwave reactor (fixed temperature, variable pressure).

Under microwave-mediated heating, the use of low absorbing solvents²⁴ such as toluene and dioxane resulted in poor conversions (entries 9–10). On the other hand, when using DMF as the solvent, although the starting material was completely consumed in 30 min, dibenzoazepinone **11** was obtained in low yield (entries 11–12). Similar results were obtained when using K₃PO₄ or KOAc as the base instead of Cs₂CO₃, and (*o*-tolyl)₃P or ^tBu₃P as the ligand (not shown in the table). Performing the reaction in acetonitrile also resulted in the complete conversion of the starting material, but **11** was again isolated only in modest yield (entry 13). It should be noted, however, that none of the experiments summarized in Table 1 yielded any observable products other than ketone **11** or the nonreacted starting material.

To compare the effectiveness of the palladium-catalyzed acylation with aldehydes with our previously reported procedure using esters as the acylating agents, we decided to prepare ester **1c** (Scheme 2). **1c** failed to undergo the acylation

Scheme 2



reaction when submitted to the conditions optimized with esters [Pd(PPh₃)₄ and K₃PO₄ in toluene at 110 °C]⁷ as well as to the new reaction conditions [Pd₂(dba)₃/ligand (dtpf or (*o*-tolyl)₃P) and Cs₂CO₃ in toluene at 110 °C]. In all the experiments only minor amounts of the hydrodehalogenation product⁷ were observed together with the unreacted starting material.

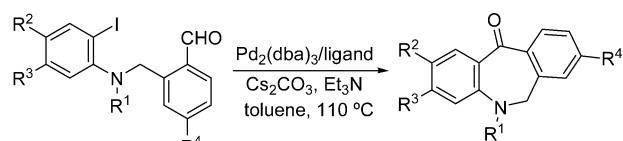
As shown in Tables 2 and 3, a variety of diversely substituted substrates was investigated under the standard thermal heating conditions using toluene as the solvent.

In general, the introduction of substituents on the aniline ring (Table 1) had little effect on the success of the acylation reaction. Thus, the nucleophilicity of the aryl palladium intermediate does not appear to be significantly affected by the electronic properties of the substituent on the aromatic ring. Interestingly, the same behavior has also been observed in the closely related palladium-catalyzed acylation reaction with esters.⁷ The functional group tolerance of the reaction is well illustrated by the fact that halides, ethers, esters, aldehydes, and free hydroxyl as well as dimethylamino groups all were perfectly accommodated. It should be noted, however, that in the acylation reactions of **4c**, minor amounts of aldehyde **14d**, resulting from the palladium-catalyzed oxidation of alcohol **14c**,¹⁷ were also isolated. The successful preparation of **14f** indicates that the Pd catalytic species are not deactivated by the presence of a strong coordinating dialkylamino group.

The effect of the substituent at the aniline nitrogen atom was also examined. While the methyl and benzyl groups were well tolerated in the acylation reaction, changing the substituent at the nitrogen from an alkyl to the electron-withdrawing *tert*-butoxycarbonyl group resulted in the formation of complex reaction mixtures in which no acylation compound could be identified (entries 22 and 23).

The palladium-catalyzed acylation was also extended to the preparation of naphtho-fused derivatives, which are attractive for their potential antitumor activity.^{20a} As can be seen in Table 3, both iodoanilines and idonaphthylamines can be used for this purpose, although the acylation from the latter proceeded more slowly.

Regarding ligands, as shown in Tables 1–3, the three sterically hindered phosphines used in this work proved to be similarly effective in promoting the acylation reaction. On the whole, dtpf²⁵ seems to be the most versatile ligand, despite not

Table 2. Synthesis of 5,6-Dihydrodibenzo[*b,e*]azepin-11-ones^a

R¹ = Bn, R² = Me, R³ = R⁴ = H, **2**
 R¹ = Me, R² = MeO, R³ = R⁴ = H, **3**
 R¹ = Me, R² = R⁴ = H, R³ = F, **4a**
 R¹ = Me, R² = R⁴ = H, R³ = Cl, **4b**
 R¹ = Me, R² = R⁴ = H, R³ = CH₂OH, **4c**
 R¹ = Me, R² = R⁴ = H, R³ = CHO, **4d**
 R¹ = Me, R² = R⁴ = H, R³ = CH₂OAc, **4e**
 R¹ = Me, R² = R⁴ = H, R³ = CH₂NMe₂, **4f**
 R¹ = Me, R² = R³ = H, R⁴ = F, **5**
 R¹ = BOC, R² = R³ = R⁴ = H, **6**

12
13
14a
14b
14c
14d
14e
14f
15

entry	aldehyde	ligand	time	yield (%) ^b
1	2	dtpf	24 h	12 (91%)
2	2	(^t Bu) ₃ P·HBF ₄	24 h	12 (77%) ^c
3	2	(<i>o</i> -tolyl) ₃ P	36 h	12 (62%)
4	3	dtpf	36 h	13 (60%)
5	3	(^t Bu) ₃ P·HBF ₄	48 h	13 (55%)
6	3	(<i>o</i> -tolyl) ₃ P	48 h	13 (51%)
7	4a	dtpf	48 h	14a (74%)
8	4a	(^t Bu) ₃ P·HBF ₄	60 h	14a (71%)
9	4b	dtpf	36 h	14b (60%) ^d
10	4b	(^t Bu) ₃ P·HBF ₄	72 h	14b (79%)
11	4b	(<i>o</i> -tolyl) ₃ P	72 h	14b (65%) ^d
12	4c	dtpf	24 h	14c (71%) ^e
13	4c	(^t Bu) ₃ P·HBF ₄	24 h	14c (53%) ^f
14	4d	dtpf	36 h	14d (53%)
15	4d	(^t Bu) ₃ P·HBF ₄	48 h	14d (46%)
16	4e	dtpf	24 h	14e (59%)
17	4e	(^t Bu) ₃ P·HBF ₄	48 h	14e (67%)
18	4f	dtpf	24 h	14f (69%)
19	5	dtpf	36 h	15 (87%)
20	5	(^t Bu) ₃ P·HBF ₄	36 h	15 (72%)
21	5	(<i>o</i> -tolyl) ₃ P	36 h	15 (55%)
22	6	dtpf	24 h	–
23	6	(<i>o</i> -tolyl) ₃ P	24 h	–

^aThe reactions were carried out using Pd₂(dba)₃ (0.05 equiv), ligand (0.1 equiv), Cs₂CO₃ (3 equiv), and Et₃N (6 equiv) in toluene at 110 °C in a sealed tube. ^bYields refer to pure products isolated by flash chromatography. ^cTraces of **2** (≤5%) were also observed in the crude reaction mixture. ^dTraces of **5** (≤5%) were also observed in the crude reaction mixture. ^e**14d** (9%) was also isolated. ^f**14d** (15%) was also isolated.

always giving the highest yield. On the other hand, the use of (*o*-tolyl)₃P as the ligand usually resulted in lower reaction rates, as reflected in the longer reaction times required for the complete consumption of the starting material.

In summary, we have demonstrated that the palladium-catalyzed intramolecular acylation of aryl iodides with aldehydes constitutes an efficient methodology for the synthesis of dihydrodibenzo[*b,e*]azepin-11-ones. The present reaction is a valuable synthetic alternative to our previously reported palladium-catalyzed acylation of aryl halides by esters and amides.

EXPERIMENTAL SECTION

General Methods. Chromatography refers to flash chromatography on silica gel. ¹H and ¹³C NMR spectra were recorded in CDCl₃

Table 3. Synthesis of Tetracyclic Ketones^a

entry	aldehyde	ligand	yield (%) ^b
1	7	dtpf ^c	16 (61%)
2	7	(^t Bu) ₃ P·HBF ₄ ^c	16 (56%)
3	8	dtpf ^c	17 (58%)
4	8	(^t Bu) ₃ P·HBF ₄	17 (33%)
5	8	(<i>o</i> -tolyl) ₃ P	17 (44%)
6	9	dtpf	18 (70%)
7	9	(^t Bu) ₃ P·HBF ₄	18 (87%)
8	9	(<i>o</i> -tolyl) ₃ P	18 (90%)
9	10	dtpf ^c	19 (64%)
10	10	(^t Bu) ₃ P·HBF ₄	19 (63%)
11	10	(<i>o</i> -tolyl) ₃ P	10 + 19 (1.5:1) ^d

^aThe reactions were carried out using Pd₂(dba)₃ (0.05 equiv), ligand (0.1 equiv), Cs₂CO₃ (3 equiv), and Et₃N (6 equiv) in toluene at 110 °C for 72 h in a sealed tube. ^bYields refer to pure products isolated by flash chromatography. ^c24 h. ^d¹H NMR ratio, yield not quantified.

solution. Chemical shifts are reported in ppm downfield (δ) from Me₄Si.

2-[N-(2-Iodophenyl)-N-methylaminomethyl]benzaldehyde (1a). A mixture of 2-iodo-N-methylaniline²⁶ (0.25 g, 1.07 mmol), K₂CO₃ (0.23 g, 1.62 mmol), and 2-(bromomethyl)benzimidazole (0.21 g, 1.07 mmol) in acetonitrile (20 mL) was stirred at reflux for 120 h; every 24 h a new portion of 2-(bromomethyl)benzimidazole (0.1 g, 0.51 mmol) was added. The mixture was concentrated, and the residue was partitioned between water and CH₂Cl₂. The organic layer was dried and concentrated. The residue was purified by chromatography (from hexanes to 97:3 hexanes–EtOAc) to give 2-[N-(2-iodophenyl)-N-methylaminomethyl]benzimidazole (315 mg, 85%).

To a cooled (0 °C) solution of 2-[N-(2-iodophenyl)-N-methylaminomethyl]benzimidazole (315 mg, 0.9 mmol) in CH₂Cl₂ (5 mL), a 1.0 M solution of DIBAL-H in hexanes (1.35 mL, 1.35 mmol) was added dropwise under argon. The resulting solution was stirred at room temperature for 3 h, cooled again, and poured into a mixture of ice and 6 N HCl. The mixture was vigorously stirred for 1 h and then extracted with CH₂Cl₂. The organic extracts were washed with 1 N NaHCO₃ aqueous solution, dried, and concentrated. The residue was purified by chromatography (from hexanes to 97:3 hexanes–EtOAc) to give **1a** (140 mg, 44%) as a yellow oil. ¹H NMR (300 MHz) δ 2.67 (s, 3H), 4.57 (s, 2H), 6.81 (ddd, *J* = 7.8, 7.2, 1.5 Hz, 1H), 7.18 (dd, *J* = 8.1, 1.5 Hz, 1H), 7.32 (ddd, *J* = 7.8, 7.2, 1.5 Hz, 1H), 7.43 (t, *J* = 7.2 Hz, 1H), 7.58 (td, *J* = 7.5, 1.5 Hz, 1H), 7.77 (d, *J* = 7.5 Hz, 1H), 7.84–7.90 (m, 2H), 10.30 (s, 1H). ¹³C NMR (75.4 MHz) δ 42.8, 57.2, 98.5,

122.4, 125.8, 127.5, 129.0, 130.4, 131.5, 133.5, 134.3, 140.1, 140.6, 153.7, 192.6. IR (NaCl) ν 1693 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{15}\text{H}_{15}\text{INO}$: 352.0193 $[\text{M} + \text{H}]^+$; found: 352.0191.

2-[N-(2-Bromophenyl)-N-methylaminomethyl]benzaldehyde (1b). A mixture of 2-bromoaniline (0.5 g, 2.9 mmol), K_2CO_3 (0.6 g, 4.35 mmol), and 2-(bromomethyl)benzotriazole (0.57 g, 2.9 mmol) in acetonitrile (40 mL) was stirred at reflux for 24 h. The mixture was concentrated, and the residue was partitioned between water and CH_2Cl_2 . The organic layer was dried and concentrated. The residue was purified by chromatography (CH_2Cl_2) to give 2-[N-(2-bromophenyl)aminomethyl]benzotriazole (0.54 g, 65%).

A mixture of 2-[N-(2-bromophenyl)aminomethyl]benzotriazole (0.54 g, 1.88 mmol), K_2CO_3 (0.52 g, 3.76 mmol), and iodomethane (1.0 mL, 16.0 mmol) in acetonitrile (5 mL) was stirred at 80 °C in a sealed tube for 2 days. The mixture was concentrated, and the residue was partitioned between water and CH_2Cl_2 . The organic layer was dried and concentrated. The residue was purified by chromatography (from hexanes to 95:5 hexanes–EtOAc) to give 2-[N-(2-bromophenyl)-N-methylaminomethyl]benzotriazole (0.39 g, 69%).

1b (140 mg, 54%) was obtained starting from 2-[N-(2-bromophenyl)-N-methylaminomethyl]benzotriazole (0.26 g, 0.86 mmol) and operating as in the preparation of **1a**. Chromatography: from hexanes to 95:5 hexanes–EtOAc. Brown solid. ^1H NMR (400 MHz) δ 2.69 (s, 3H), 4.60 (s, 2H), 6.92 (ddd, $J = 8.4, 7.6, 1.6$ Hz, 1H), 7.16 (dd, $J = 8.0, 1.6$ Hz, 1H), 7.26 (m, 1H), 7.43 (t, $J = 8.0$ Hz, 1H), 7.57 (d, $J = 8.0$ Hz, 1H), 7.58 (t, $J = 7.6$ Hz, 1H), 7.78 (d, $J = 7.6$ Hz, 1H), 7.86 (d, $J = 8.0$ Hz, 1H), 10.27 (s, 1H). ^{13}C NMR (100.5 MHz) δ 41.5, 57.0, 120.3, 122.3, 124.7, 127.4, 128.1, 129.9, 131.8, 133.6, 133.9, 134.3, 140.9, 150.9, 192.6. IR (NaCl) ν 1693 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{15}\text{H}_{15}\text{BrNO}$: 304.0332 $[\text{M} + \text{H}]^+$; found: 304.0333.

Compounds **1c** and **2** were prepared according to the preparation of **1a**.

Methyl 2-[N-(2-iodophenyl)-N-methylaminomethyl]benzoate (1c). Chromatography: from hexanes to 95:5 hexanes–EtOAc. Pale yellow oil. ^1H NMR (300 MHz) δ 2.64 (s, 3H), 3.84 (s, 3H), 4.53 (s, 2H), 6.78 (ddd, $J = 8.1, 7.5, 1.5$ Hz, 1H), 7.15 (dd, $J = 8.1, 1.5$ Hz, 1H), 7.25–7.34 (m, 2H), 7.51 (td, $J = 7.5, 1.5$ Hz, 1H), 7.83–7.91 (m, 3H). ^{13}C NMR (75.4 MHz) δ 42.5, 52.0, 58.4, 98.1, 122.2, 125.3, 126.6, 129.0, 129.7, 130.1, 130.4, 131.8, 140.0, 140.1, 154.4, 168.0. HRMS (ESI-TOF) calcd for $\text{C}_{16}\text{H}_{17}\text{INO}_2$: 382.0304 $[\text{M} + \text{H}]^+$; found: 382.0306.

2-[N-Benzyl-N-(2-iodo-4-methylphenyl)aminomethyl]benzaldehyde (2). Chromatography: from hexanes to 9:1 hexanes–EtOAc. Colorless oil. ^1H NMR (300 MHz) δ 2.21 (s, 3H), 4.11 (s, 2H), 4.49 (s, 2H), 6.83 (d, $J = 7.8$ Hz, 1H), 6.98 (dd, $J = 7.8, 2.1$ Hz, 1H), 7.21–7.41 (m, 6H), 7.48 (td, $J = 7.5, 1.5$ Hz, 2H), 7.59 (d, $J = 7.8$ Hz, 1H), 7.67 (dd, $J = 2.1, 0.6$ Hz, 1H), 7.80 (dd, $J = 7.8, 1.5$ Hz, 1H), 10.13 (s, 1H). ^{13}C NMR (75.4 MHz) δ 20.2, 52.8, 59.0, 99.9, 124.2, 127.3, 127.4, 128.2, 129.1, 129.4, 130.8, 130.9, 133.3, 134.4, 136.2, 137.3, 140.3, 140.4, 148.2, 192.3. IR (NaCl) ν 1693 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{22}\text{H}_{21}\text{INO}$: 442.0662 $[\text{M} + \text{H}]^+$; found: 442.0655.

Compounds **3**, **4a–c**, **5**, and **8–10** were prepared according to the preparation of **1b** starting from the corresponding *o*-iodoaryl amines.^{27–30}

2-[N-(2-Iodo-4-methoxyphenyl)-N-methylaminomethyl]benzaldehyde (3). Chromatography: CH_2Cl_2 . Brown solid. ^1H NMR (300 MHz) δ 2.59 (s, 3H), 3.75 (s, 3H), 4.47 (s, 2H), 6.87 (dd, $J = 8.7, 3.0$ Hz, 1H), 7.10 (d, $J = 8.7$ Hz, 1H), 7.37 (d, $J = 3.0$ Hz, 1H), 7.41 (td, $J = 7.5, 0.9$ Hz, 1H), 7.54 (td, $J = 7.5, 1.5$ Hz, 1H), 7.70 (d, $J = 7.5$ Hz, 1H), 7.87 (dd, $J = 7.5, 1.2$ Hz, 1H), 10.34 (s, 1H). ^{13}C NMR (75.4 MHz) δ 43.4, 55.6, 57.5, 99.7, 114.8, 122.7, 124.6, 127.5, 130.6, 131.0, 133.4, 134.3, 140.7, 146.7, 158.7, 192.6. IR (NaCl) ν 1693 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{16}\text{H}_{17}\text{INO}_2$: 382.0298 $[\text{M} + \text{H}]^+$; found: 382.0290.

2-[N-(5-Fluoro-2-iodophenyl)-N-methylaminomethyl]benzaldehyde (4a). Chromatography: CH_2Cl_2 . Yellow oil. ^1H NMR (300 MHz) δ 2.67 (s, 3H), 4.56 (s, 2H), 6.59 (ddd, $J = 8.7, 7.8, 3.0$ Hz, 1H), 6.90 (dd, $J = 10.5, 3.0$ Hz, 1H), 7.45 (td, $J = 7.5, 1.2$ Hz, 1H),

7.59 (td, $J = 7.5, 1.5$ Hz, 1H), 7.77 (dd, $J = 8.7, 6.3$ Hz, 1H), 7.78 (d, $J = 7.8$ Hz, 1H), 7.87 (dd, $J = 7.5, 1.5$ Hz, 1H), 10.27 (s, 1H). ^{13}C NMR (75.4 MHz) δ 42.5, 57.2, 90.3 (d, $J = 4.1$ Hz), 109.9 (d, $J = 23.0$ Hz), 112.8 (d, $J = 21.9$ Hz), 127.6, 130.2, 132.2, 133.6, 134.2, 140.1, 140.6 (d, $J = 9.2$ Hz), 155.5 (d, $J = 8.1$ Hz), 163.6 (d, $J = 248.0$ Hz), 192.7. IR (NaCl) ν 1692 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{15}\text{H}_{14}\text{FINO}$: 370.0099 $[\text{M} + \text{H}]^+$; found: 370.0110.

2-[N-(5-Chloro-2-iodophenyl)-N-methylaminomethyl]benzaldehyde (4b). Chromatography: from hexanes to 95:5 hexanes–EtOAc. Yellow oil. ^1H NMR (300 MHz) δ 2.66 (s, 3H), 4.55 (s, 2H), 6.81 (dd, $J = 8.1, 2.4$ Hz, 1H), 7.14 (d, $J = 2.4$ Hz, 1H), 7.47 (td, $J = 7.8, 1.2$ Hz, 1H), 7.59 (td, $J = 7.8, 1.5$ Hz, 1H), 7.75 (d, $J = 8.1$ Hz, 1H), 7.78 (d, $J = 7.8$ Hz, 1H), 7.87 (dd, $J = 7.8, 1.5$ Hz, 1H), 10.27 (s, 1H). ^{13}C NMR (75.4 MHz) δ 42.5, 57.2, 95.0, 122.8, 125.7, 127.6, 130.2, 132.2, 133.6, 134.2, 134.9, 140.2, 140.7, 155.1, 192.6. IR (NaCl) ν 1699 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{15}\text{H}_{14}\text{ClINO}$: 385.9803 $[\text{M} + \text{H}]^+$; found: 385.9797.

2-[N-(5-Hydroxymethyl-2-iodophenyl)-N-methylaminomethyl]benzaldehyde (4c). Chromatography: from CH_2Cl_2 to CH_2Cl_2 –MeOH 1%. Colorless oil. ^1H NMR (400 MHz) δ 1.98 (broad s, 1H), 2.65 (s, 3H), 4.55 (s, 2H), 4.63 (s, 2H), 6.81 (dd, $J = 8.0, 2.0$ Hz, 1H), 7.19 (d, $J = 2.0$ Hz, 1H), 7.43 (td, $J = 7.6, 0.8$ Hz, 1H), 7.58 (td, $J = 7.6, 1.6$ Hz, 1H), 7.77 (d, $J = 7.6$ Hz, 1H), 7.81 (d, $J = 8.0$ Hz, 1H), 7.86 (dd, $J = 7.6, 1.6$ Hz, 1H), 10.25 (s, 1H). ^{13}C NMR (100.5 MHz) δ 42.6, 57.2, 64.6, 96.9, 120.9, 124.3, 127.6, 130.5, 131.5, 133.5, 134.3, 140.1, 140.6, 142.3, 153.9, 192.6. IR (NaCl) ν 1682, 3400 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{16}\text{H}_{17}\text{INO}_2$: 382.0298 $[\text{M} + \text{H}]^+$; found: 382.0298.

2-[N-(5-Formyl-2-iodophenyl)-N-methylaminomethyl]benzaldehyde (4d). **4d** was prepared from **4c** (Swern oxidation). Chromatography: CH_2Cl_2 . Yellow oil. ^1H NMR (400 MHz) δ 2.72 (s, 3H), 4.63 (s, 2H), 7.28 (dd, $J = 8.4, 2.0$ Hz, 1H), 7.46 (td, $J = 7.6, 0.8$ Hz, 1H), 7.58 (td, $J = 7.6, 1.6$ Hz, 1H), 7.65 (d, $J = 2.0$ Hz, 1H), 7.77 (d, $J = 8.0$ Hz, 1H), 7.88 (dd, $J = 7.6, 1.6$ Hz, 1H), 8.06 (d, $J = 8.0$ Hz, 1H), 9.95 (s, 1H), 10.29 (s, 1H). ^{13}C NMR (100.5 MHz) δ 42.5, 57.3, 106.7, 121.5, 126.8, 127.7, 130.3, 132.1, 133.6, 134.2, 137.2, 140.0, 141.2, 154.9, 191.3, 192.6. IR (NaCl) ν 1681, 1704 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{16}\text{H}_{15}\text{INO}_2$: 380.0142 $[\text{M} + \text{H}]^+$; found: 380.0135.

2-[N-(5-Acetoxyethyl-2-iodophenyl)-N-methylaminomethyl]benzaldehyde (4e). **4e** was prepared from **4c** (AcCl). Chromatography: CH_2Cl_2 . Pale yellow oil. ^1H NMR (400 MHz) δ 2.11 (s, 3H), 2.67 (s, 3H), 4.56 (s, 2H), 5.03 (s, 2H), 6.81 (dd, $J = 8.0, 1.6$ Hz, 1H), 7.15 (d, $J = 1.6$ Hz, 1H), 7.43 (t, $J = 7.6$ Hz, 1H), 7.57 (td, $J = 7.6, 1.6$ Hz, 1H), 7.77 (d, $J = 7.6$ Hz, 1H), 7.82 (d, $J = 8.0$ Hz, 1H), 7.87 (dd, $J = 7.6, 1.2$ Hz, 1H), 10.28 (s, 1H). ^{13}C NMR (100.5 MHz) δ 20.9, 42.6, 57.2, 65.5, 97.8, 122.1, 125.5, 127.5, 130.4, 131.6, 133.5, 134.3, 137.2, 140.3, 140.4, 153.9, 170.6, 192.6. IR (NaCl) ν 1693, 1738 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{18}\text{H}_{19}\text{INO}_3$: 424.0404 $[\text{M} + \text{H}]^+$; found: 424.0394.

2-[N-(5-Dimethylaminomethyl-2-iodophenyl)-N-methylaminomethyl]benzaldehyde (4f). **4f** was prepared from **4c** (a. MsCl, b. Me_2NH). Chromatography: from CH_2Cl_2 to CH_2Cl_2 –MeOH 8%. Brown oil. ^1H NMR (400 MHz) δ 2.29 (s, 6H), 2.68 (s, 3H), 3.46 (s, 2H), 4.58 (s, 2H), 6.77 (dd, $J = 8.0, 2.0$ Hz, 1H), 7.22 (s, 1H), 7.42 (t, $J = 7.6$ Hz, 1H), 7.55 (td, $J = 7.6, 1.6$ Hz, 1H), 7.75 (d, $J = 7.6$ Hz, 1H), 7.79 (d, $J = 8.0$ Hz, 1H), 7.87 (dd, $J = 7.6, 1.6$ Hz, 1H), 10.32 (s, 1H). ^{13}C NMR (100.5 MHz) δ 42.8, 45.2, 57.1, 63.6, 96.5, 123.0, 126.5, 127.5, 130.5, 131.1, 133.4, 134.3, 139.8, 140.0, 140.7, 153.6, 192.5. IR (NaCl) ν 1693 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{18}\text{H}_{22}\text{IN}_2\text{O}$: 409.0771 $[\text{M} + \text{H}]^+$; found: 409.0774.

4-Fluoro-2-[N-(2-iodophenyl)-N-methylaminomethyl]benzaldehyde (5). Chromatography: CH_2Cl_2 . Green oil. ^1H NMR (400 MHz) δ 2.69 (s, 3H), 4.57 (s, 2H), 6.82 (ddd, $J = 8, 7.2, 1.6$ Hz, 1H), 7.10 (td, $J = 8, 2.8$ Hz, 1H), 7.19 (dd, $J = 8, 1.6$ Hz, 1H), 7.32 (ddd, $J = 8, 7.2, 1.6$ Hz, 1H), 7.66 (dd, $J = 10.4, 2.4$ Hz, 1H), 7.87 (m, 2H), 10.18 (s, 1H). ^{13}C NMR (100.5 MHz) δ 42.7, 57.2 (d, $J = 1.5$ Hz), 98.4, 114.4 (d, $J = 21.4$ Hz), 117.4 (d, $J = 22.9$ Hz), 122.4, 126.0, 129.2, 130.6 (d, $J = 2.3$ Hz), 135.2 (d, $J = 9.9$ Hz), 140.2, 144.8 (d, $J = 8.4$ Hz), 153.6, 166.0 (d, $J = 255.8$ Hz), 191.1. IR (NaCl) ν 1694 cm^{-1} .

HRMS (ESI-TOF) calcd for $C_{15}H_{14}FNO$: 370.0099 $[M + H]^+$; found: 370.0096.

2-[*N*-(*tert*-Butoxycarbonyl)-*N*-(2-iodophenyl)aminomethyl]-benzaldehyde (6). A solution of *N*-(*tert*-butoxycarbonyl)-2-iodoaniline (0.4 g, 1.25 mmol) in THF (2 mL) was added at 0 °C under argon to a suspension of NaH (60% in oil, 65 mg, 1.63 mmol) in THF (8 mL). After 15 min at 25 °C, the mixture was cooled again to 0 °C. DMF (2 mL) and 2-(bromomethyl)benzaldehyde (325 mg, 1.63 mmol) were added and the mixture was stirred for 10 min at 0 °C and at 25 °C overnight. Saturated aqueous NH_4Cl was added, and the mixture was extracted with EtOAc. The organic extracts were dried and concentrated. The residue was purified by chromatography (from hexanes to 8:2 hexanes–EtOAc) to give **6** (265 mg, 48%) as an orange oil. 1H NMR (400 MHz, mixture of rotamers) δ 1.38 (s, 9H), 4.91 (d, $J = 16$ Hz, 1H), 5.48 (d, $J = 16$ Hz, 1H), 6.77–7.84 (m, 8H), 10.12 (s, 1H). ^{13}C NMR (100.5 MHz, major rotamer) δ 28.2, 48.6, 80.8, 100.3, 127.9, 128.6, 128.8, 129.8, 130.7, 130.9, 133.7, 134.3, 139.6, 143.5, 153.9, 192.1. HRMS (ESI-TOF) calcd for $C_{19}H_{21}INO_3$: 438.0561 $[M + H]^+$; found: 438.0549.

3-[*N*-(2-Iodophenyl)-*N*-methylaminomethyl]-2-naphthaldehyde (7). A mixture of 2-iodo-*N*-methylaniline (150 g, 0.68 mmol), 3-(bromomethyl)-2-naphthaldehyde³¹ (115 mg, 0.46 mmol), and 2,6-lutidine (80 μ L, 0.68 mmol) was stirred at 100 °C for 6 h. The reaction mixture was partitioned between water and Et₂O. The organic layer was dried and concentrated. The residue was purified by chromatography (from hexanes to 95:5 hexanes–EtOAc) to give **7** (130 mg, 70%) as an orange oil. 1H NMR (300 MHz) δ 2.72 (s, 3H), 4.70 (s, 2H), 6.80 (ddd, $J = 7.8, 7.2, 1.5$ Hz, 1H), 7.25 (dd, $J = 7.8, 1.5$ Hz, 1H), 7.33 (ddd, $J = 7.8, 7.2, 1.5$ Hz, 1H), 7.54 (td, $J = 7.2, 1.2$ Hz, 1H), 7.62 (td, $J = 7.2, 1.2$ Hz, 1H), 7.86 (dd, $J = 7.8, 1.5$ Hz, 1H), 7.90 (d, $J = 7.8$ Hz, 1H), 7.97 (d, $J = 7.8$ Hz, 1H), 8.17 (s, 1H), 8.39 (s, 1H), 10.36 (s, 1H). ^{13}C NMR (75.4 MHz) δ 43.0, 57.8, 98.5, 122.4, 125.7, 126.7, 127.8, 129.1 (2 CH), 129.2, 129.4, 131.7, 132.7, 135.2, 135.3, 135.5, 140.1, 154.0, 192.9. IR (NaCl) ν 1692 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{19}H_{17}INO$: 402.0349 $[M + H]^+$; found: 402.0355.

2-[*N*-(3-Iodo-2-naphthyl)-*N*-methylaminomethyl]-benzaldehyde (8). Chromatography: from hexanes to 95:5 hexanes–EtOAc. Yellow solid. 1H NMR (300 MHz) δ 2.73 (s, 3H), 4.66 (s, 2H), 7.35–7.48 (m, 3H), 7.52 (s, 1H), 7.57 (ddd, $J = 7.8, 7.2, 1.5$ Hz, 1H), 7.65 (d, $J = 7.8$ Hz, 1H), 7.72 (d, $J = 8.1$ Hz, 1H), 7.83 (d, $J = 7.8$ Hz, 1H), 7.87 (dd, $J = 7.8, 1.2$ Hz, 1H), 8.40 (s, 1H), 10.32 (s, 1H). ^{13}C NMR (75.4 MHz) δ 43.3, 57.5, 98.0, 119.0, 125.4, 126.4, 126.7, 127.0, 127.5, 130.4, 131.7, 132.1, 133.5, 133.6, 134.3, 139.7, 140.6, 150.5, 192.7. IR (NaCl) ν 1691 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{19}H_{17}INO$: 402.0349 $[M + H]^+$; found: 402.0342.

2-[*N*-(1-Iodo-2-naphthyl)-*N*-methylaminomethyl]-benzaldehyde (9). Chromatography: from hexanes to 1:1 hexanes– CH_2Cl_2 . Yellow oil. 1H NMR (400 MHz) δ 2.79 (s, 3H), 4.69 (s, 2H), 7.40–7.48 (m, 3H), 7.52–7.58 (m, 2H), 7.75 (dd, $J = 7.6, 0.8$ Hz, 1H), 7.81 (d, $J = 8.0$ Hz, 1H), 7.82 (d, $J = 7.6$ Hz, 1H), 7.89 (dd, $J = 7.6, 1.2$ Hz, 1H), 8.26 (d, $J = 8.4$ Hz, 1H), 10.40 (s, 1H). ^{13}C NMR (100.5 MHz) δ 43.6, 57.4, 103.5, 121.4, 125.7, 127.6, 127.8, 128.0, 129.8, 130.7, 131.5, 132.0, 132.9, 133.6, 134.3, 135.9, 140.7, 152.3, 192.8. IR (NaCl) ν 1692 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{19}H_{17}INO$: 402.0349 $[M + H]^+$; found: 402.0350.

2-[*N*-(2-Iodo-1-naphthyl)-*N*-methylaminomethyl]-benzaldehyde (10). Chromatography: from hexanes to 9:1 hexanes–EtOAc. Yellow oil. 1H NMR (300 MHz) δ 3.02 (s, 3H), 4.91 (d, $J = 13.5$ Hz, 1H), 4.97 (d, $J = 13.5$ Hz, 1H), 7.39 (d, $J = 8.4$ Hz, 1H), 7.41–7.51 (m, 3H), 7.54 (td, $J = 7.5, 1.5$ Hz, 1H), 7.63 (d, $J = 7.5$ Hz, 1H), 7.82 (m, 1H), 7.85 (d, $J = 9$ Hz, 1H), 7.87 (m, 1H), 8.05 (m, 1H), 20.21 (s, 1H). ^{13}C NMR (100.5 MHz) δ 40.6, 56.2, 97.3, 124.0, 126.2, 126.4, 127.5, 127.6, 128.5, 130.8, 131.2, 133.4, 133.6, 134.3, 134.6, 136.5, 141.6, 149.5, 192.1. IR (NaCl) ν 1691 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{19}H_{17}INO$: 402.0349 $[M + H]^+$; found: 402.0340.

Representative Procedure for the Pd(0)-Catalyzed Acylation (Table 1, entry 1). A mixture of aldehyde **1a** (70 mg, 0.2 mmol), CS_2CO_3 (195 mg, 0.6 mmol), Et₃N (0.17 mL, 1.2 mmol), Pd₂(dba)₃

(9 mg, 0.01 mmol), and dtpf (9.5 mg, 0.02 mmol) in toluene (8 mL) was stirred at 110 °C in a sealed tube for 24 h. The reaction mixture was partitioned between water and Et₂O. The organic extracts were washed with brine, dried, and concentrated. The residue was purified by chromatography (from hexanes to hexanes–EtOAc 2.5%) to give **11** (34 mg, 77%).

6-Methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (11). Yellow solid: mp 109–110 °C (recrystallized from Et₂O). 1H NMR (300 MHz) δ 3.24 (s, 3H), 4.26 (s, 2H), 6.85 (m, 1H), 6.87 (d, $J = 7.8$ Hz, 1H), 7.24 (dd, $J = 7.2, 1.5$ Hz, 1H), 7.34–7.43 (m, 2H), 7.49 (td, $J = 7.5, 1.5$ Hz, 1H), 7.76 (dd, $J = 7.5, 1.5$ Hz, 1H), 8.30 (dd, $J = 8.4, 1.5$ Hz, 1H). ^{13}C NMR (75.4 MHz) δ 40.1, 58.4, 115.7, 117.5, 123.5, 125.6, 128.0, 128.9, 131.9, 132.3, 134.1, 136.9, 140.5, 150.9, 193.4. IR (NaCl) ν 1629 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{15}H_{14}NO$: 224.1070 $[M + H]^+$; found: 224.1069.

6-Benzyl-9-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (12). Chromatography: from hexanes to hexanes–EtOAc 3%. Yellow solid: mp 85–90 °C (recrystallized from Et₂O). Yield: 91% (48 mg). 1H NMR (300 MHz) δ 2.30 (s, 3H), 4.30 (s, 2H), 4.70 (s, 2H), 6.80 (d, $J = 8.4$ Hz, 1H), 7.08 (dd, $J = 7.5, 1.2$ Hz, 1H), 7.15 (dd, $J = 8.4, 2.4$ Hz, 1H), 7.22 (d, $J = 7.2$ Hz, 2H), 7.30–7.43 (m, 4H), 7.46 (td, $J = 7.5, 1.5$ Hz, 1H), 7.78 (dd, $J = 7.5, 1.2$ Hz, 1H), 8.12 (d, $J = 1.2$ Hz, 1H). ^{13}C NMR (75.4 MHz) δ 20.2, 55.4, 55.9, 117.2, 123.5, 125.9, 126.7, 127.3, 127.4, 128.0, 128.8, 129.0, 131.5, 131.8, 135.6, 136.7, 137.6, 140.7, 149.2, 193.6. IR (NaCl) ν 1633 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{22}H_{20}NO$: 314.1545 $[M + H]^+$; found: 314.1548.

9-Methoxy-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (13). Chromatography: from hexanes to hexanes–EtOAc 5%. Brown solid: mp 95–97 °C (recrystallized from Et₂O). Yield: 60% (30 mg). 1H NMR (300 MHz) δ 3.19 (s, 3H), 3.84 (s, 3H), 4.23 (s, 2H), 6.88 (d, $J = 9.0$ Hz, 1H), 7.08 (dd, $J = 9.0, 3.3$ Hz, 1H), 7.24 (d, $J = 7.8$ Hz, 1H), 7.38 (ddd, $J = 7.8, 7.5, 1.2$ Hz, 1H), 7.50 (dd, $J = 7.8, 7.5, 1.5$ Hz, 1H), 7.78 (d, $J = 3.3$ Hz, 1H), 7.79 (d, $J = 7.8$ Hz, 1H). ^{13}C NMR (75.4 MHz) δ 40.2, 55.6, 58.5, 112.1, 118.2, 123.7, 124.3, 125.8, 127.9, 129.1, 131.9, 137.0, 139.9, 146.5, 151.8, 192.7. IR (NaCl) ν 1630 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{16}H_{16}NO_2$: 254.1176 $[M + H]^+$; found: 254.1182.

8-Fluoro-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (14a). Chromatography: from hexanes to hexanes–EtOAc 10%. Yellow gum. Yield: 74% (36 mg). 1H NMR (300 MHz) δ 3.23 (s, 3H), 4.28 (s, 2H), 6.51 (dd, $J = 12.0, 2.4$ Hz, 1H), 6.90 (ddd, $J = 9.6, 7.5, 2.4$ Hz, 1H), 7.25 (d, $J = 7.5$ Hz, 1H), 7.39 (td, $J = 7.5, 1.2$ Hz, 1H), 7.51 (td, $J = 7.5, 1.5$ Hz, 1H), 7.76 (dd, $J = 7.5, 1.2$ Hz, 1H), 8.32 (dd, $J = 9.0, 7.5$ Hz, 1H). ^{13}C NMR (75.4 MHz) δ 40.3, 58.5, 101.2 (d, $J = 25.9$ Hz), 106.0 (d, $J = 23.0$ Hz), 120.4, 125.7, 128.3, 129.0, 132.0, 135.4 (d, $J = 11.5$ Hz), 136.5, 140.3, 152.5 (d, $J = 12.1$ Hz), 166.7 (d, $J = 25.2$ S Hz), 192.0. IR (NaCl) ν 1633 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{15}H_{13}FNO$: 242.0976 $[M + H]^+$; found: 242.0981.

8-Chloro-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (14b). Chromatography: from hexanes to hexanes–EtOAc 8%. Brown solid: mp 86–88 °C. Yield: 79% (35 mg). 1H NMR (400 MHz) δ 3.24 (s, 3H), 4.26 (s, 2H), 6.82 (dd, $J = 8.8, 2.0$ Hz, 1H), 6.85 (s, 1H), 7.25 (d, $J = 7.6$ Hz, 1H), 7.39 (dd, $J = 8.0, 7.2$ Hz, 1H), 7.51 (ddd, $J = 7.6, 7.2, 1.2$ Hz, 1H), 7.75 (d, $J = 8.0$ Hz, 1H), 8.23 (d, $J = 8.8$ Hz, 1H). ^{13}C NMR (100.5 MHz) δ 40.2, 58.4, 115.1, 118.1, 122.0, 125.8, 128.3, 129.0, 132.2, 134.0, 136.6, 140.2, 140.3, 151.3, 192.3. IR (NaCl) ν 1630 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{15}H_{13}ClNO$: 258.0680 $[M + H]^+$; found: 258.0678.

8-(Hydroxymethyl)-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (14c). Chromatography: from CH_2Cl_2 to CH_2Cl_2 –MeOH 1%. Yellow oil. Yield: 71% (32 mg). 1H NMR (400 MHz) δ 2.39 (broad s, 1H), 3.22 (s, 3H), 4.20 (s, 2H), 4.67 (s, 2H), 6.77 (dd, $J = 8.0, 1.6$ Hz, 1H), 6.89 (s, 1H), 7.22 (d, $J = 7.6$ Hz, 1H), 7.36 (td, $J = 7.6, 1.2$ Hz, 1H), 7.48 (td, $J = 7.6, 1.2$ Hz, 1H), 7.74 (dd, $J = 7.2, 1.2$ Hz, 1H), 8.24 (d, $J = 8.0$ Hz, 1H). ^{13}C NMR (100.5 MHz) δ 40.1, 58.4, 64.7, 113.0, 115.8, 122.5, 125.7, 128.0, 128.8, 132.0, 132.7, 136.8, 140.3, 147.6, 151.1, 193.1. IR (NaCl) ν 1623, 3406 cm^{-1} . HRMS (ESI-TOF) calcd for $C_{16}H_{16}NO_2$: 254.1176 $[M + H]^+$; found: 254.1166.

8-(Formyl)-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (14d). Chromatography: from hexanes to CH_2Cl_2 . Yellow oil. Yield:

49% (22 mg). ^1H NMR (400 MHz) δ 3.35 (s, 3H), 4.32 (s, 2H), 7.28 (d, $J = 7.6$ Hz, 1H), 7.32 (dd, $J = 8.4, 1.6$ Hz, 1H), 7.40 (s, 1H), 7.41 (td, $J = 7.6, 1.2$ Hz, 1H), 7.55 (td, $J = 7.6, 1.6$ Hz, 1H), 7.75 (dd, $J = 7.6, 1.2$ Hz, 1H), 8.43 (d, $J = 8.0$ Hz, 1H). ^{13}C NMR (100.5 MHz) δ 40.3, 58.4, 117.2, 117.4, 125.9, 126.8, 128.4, 129.0, 132.5, 133.5, 136.7, 139.8, 140.2, 150.8, 192.4, 193.3. IR (NaCl) ν 1633, 1693 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{16}\text{H}_{14}\text{NO}_2$: 252.1019 $[\text{M} + \text{H}]^+$; found: 252.1016.

8-(Acetoxymethyl)-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (14e). Chromatography: from hexanes to CH_2Cl_2 . Yellow oil. Yield: 69% (32 mg). ^1H NMR (400 MHz) δ 2.12 (s, 3H), 3.26 (s, 3H), 4.26 (s, 2H), 5.08 (s, 2H), 6.83 (d, $J = 7.6$ Hz, 1H), 6.85 (s, 1H), 7.24 (d, $J = 7.2$ Hz, 1H), 7.38 (td, $J = 7.6, 1.2$ Hz, 1H), 7.50 (td, $J = 7.6, 1.2$ Hz, 1H), 7.75 (dd, $J = 7.2, 1.2$ Hz, 1H), 8.29 (d, $J = 8.0$ Hz, 1H). ^{13}C NMR (100.5 MHz) δ 20.9, 40.1, 58.4, 65.8, 114.8, 116.9, 123.1, 125.7, 128.1, 128.9, 132.0, 132.9, 136.8, 140.3, 142.1, 150.8, 170.7, 192.9. IR (NaCl) ν 1632, 1738 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{18}\text{H}_{18}\text{NO}_3$: 296.1281 $[\text{M} + \text{H}]^+$; found: 296.1276.

8-(Dimethylaminomethyl)-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (14f). Chromatography (SiO_2 , from CH_2Cl_2 to CH_2Cl_2 -MeOH 3%). Yellow oil. Yield: 69% (28 mg). ^1H NMR (400 MHz) δ 2.27 (s, 6H), 3.27 (s, 3H), 3.44 (s, 2H), 4.26 (s, 2H), 6.81 (dd, $J = 8.4, 1.6$ Hz, 1H), 6.89 (s, 1H), 7.25 (d, $J = 7.6$ Hz, 1H), 7.38 (td, $J = 7.6, 1.2$ Hz, 1H), 7.49 (td, $J = 7.6, 1.2$ Hz, 1H), 7.76 (dd, $J = 7.6, 1.2$ Hz, 1H), 8.25 (d, $J = 8.4$ Hz, 1H). ^{13}C NMR (100.5 MHz) δ 40.2, 45.4, 58.4, 64.2, 115.8, 118.6, 122.7, 125.7, 128.0, 128.9, 131.9, 132.5, 136.9, 140.5, 145.1, 151.0, 193.0. IR (NaCl) ν 1633 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{18}\text{H}_{21}\text{N}_2\text{O}$: 281.1654 $[\text{M} + \text{H}]^+$; found: 281.1654.

4-Fluoro-6-methyl-5,6-dihydrodibenzo[*b,e*]azepin-11-one (15). Chromatography: from hexanes to hexanes- CH_2Cl_2 80%. Yellow solid: mp 82–84 $^\circ\text{C}$. Yield: 87% (34 mg). ^1H NMR (400 MHz) δ 3.24 (s, 3H), 4.23 (s, 2H), 6.88 (m, 2H), 6.95 (dd, $J = 8.8, 2.4$ Hz, 1H), 7.07 (td, $J = 8.8, 2.4$ Hz, 1H), 7.41 (ddd, $J = 8.8, 7.2, 1.2$ Hz, 1H), 7.79 (dd, $J = 8.8, 5.6$ Hz, 1H), 8.31 (dd, $J = 8.8, 1.2$ Hz, 1H). ^{13}C NMR (100.5 MHz) δ 40.2, 58.0 (d, $J = 1.5$ Hz), 112.6 (d, $J = 21.8$ Hz), 115.1 (d, $J = 21.0$ Hz), 115.9, 117.9, 123.5, 131.7 (d, $J = 9.2$ Hz), 132.4, 134.2, 136.7 (d, $J = 3.1$ Hz), 139.3 (d, $J = 7.0$ Hz), 150.9, 165.0 (d, $J = 252.8$ Hz), 191.6. IR (NaCl) ν 1633 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{15}\text{H}_{13}\text{FNO}$: 242.0975 $[\text{M} + \text{H}]^+$; found: 242.0976.

5-Methyl-5,6-dihydrobenzo[*b*]naphtho[2,3-*e*]azepin-13-one (16). Chromatography: from hexanes to hexanes-EtOAc 8%. Yellow oil. Yield: 61% (21 mg). ^1H NMR (300 MHz) δ 3.25 (s, 3H), 4.43 (s, 2H), 6.84–6.91 (m, 2H), 7.40 (ddd, $J = 8.7, 6.9, 1.5$ Hz, 1H), 7.46–7.58 (m, 2H), 7.67 (s, 1H), 7.86 (d, $J = 8.1$ Hz, 1H), 7.92 (d, $J = 8.1$ Hz, 1H), 8.29 (s, 1H), 8.35 (dd, $J = 8.7, 1.5$ Hz, 1H). ^{13}C NMR (75.4 MHz) δ 39.9, 58.8, 115.8, 117.6, 123.7, 123.9, 126.3, 127.3, 127.8, 129.2, 129.7, 132.3, 132.6, 134.1, 134.4, 134.9, 139.0, 151.2, 193.4. IR (NaCl) ν 1619, 1639 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{19}\text{H}_{16}\text{NO}$: 274.1226 $[\text{M} + \text{H}]^+$; found: 274.1223.

6-Methyl-6,7-dihydrobenzo[*e*]naphtho[2,3-*b*]azepin-12-one (17). Chromatography: from hexanes to 1:1 hexanes- CH_2Cl_2 . Red oil. Yield: 58% (30 mg). ^1H NMR (300 MHz) δ 3.28 (s, 3H), 4.30 (s, 2H), 7.08 (s, 1H), 7.22 (ddd, $J = 8.4, 7.2, 1.2$ Hz, 1H), 7.27 (dd, $J = 7.5, 0.9$ Hz, 1H), 7.36–7.45 (m, 2H), 7.51 (td, $J = 7.5, 1.5$ Hz, 1H), 7.62 (d, $J = 8.4$ Hz, 1H), 7.83 (d, $J = 7.5$ Hz, 1H), 7.84 (dd, $J = 7.5, 3.3$ Hz, 1H), 8.29 (s, 1H). ^{13}C NMR (75.4 MHz) δ 40.9, 58.5, 109.7, 123.0, 125.7, 125.9, 126.6, 126.8, 128.0, 128.8, 129.2, 129.8, 132.2, 134.5, 137.3, 138.2, 139.8, 147.4, 194.0. IR (NaCl) ν 1619, 1643 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{19}\text{H}_{16}\text{NO}$: 274.1226 $[\text{M} + \text{H}]^+$; found: 274.1224.

7-Methyl-7,8-dihydrobenzo[*e*]naphtho[2,1-*b*]azepin-13-one (18). Chromatography: from hexanes to 1:1 hexanes-EtOAc. Yellow oil. Yield: 90% (46 mg). ^1H NMR (400 MHz) δ 3.29 (s, 3H), 4.32 (s, 2H), 7.10 (d, $J = 8.8$ Hz, 1H), 7.24 (d, $J = 7.2$ Hz, 1H), 7.32 (dd, $J = 8.4, 7.2, 1.2$ Hz, 1H), 7.37 (td, $J = 7.6, 1.2$ Hz, 1H), 7.46 (td, $J = 7.6, 1.6$ Hz, 1H), 7.58 (ddd, $J = 8.8, 7.2, 1.2$ Hz, 1H), 7.65 (dd, $J = 8.0, 1.2$ Hz, 1H), 7.72 (d, $J = 9.2$ Hz, 1H), 7.82 (dd, $J = 8.0, 1.2$ Hz, 1H), 9.32 (d, $J = 8.4$ Hz, 1H). ^{13}C NMR (100.5 MHz) δ 39.7, 58.0, 114.4, 117.0, 123.4, 124.8, 126.9, 127.7, 128.1, 128.2, 128.6, 129.0, 130.8, 134.1,

135.1, 137.1, 142.1, 152.4, 191.9. IR (NaCl) ν 1603, 1635 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{19}\text{H}_{16}\text{NO}$: 274.1226 $[\text{M} + \text{H}]^+$; found: 274.1228.

13-Methyl-12,13-dihydrobenzo[*e*]naphtho[1,2-*b*]azepin-7-one (19). Chromatography: from hexanes to 1:1 hexanes- CH_2Cl_2 . Yellow oil. Yield: 64% (30 mg). ^1H NMR (400 MHz) δ 2.93 (s, 3H), 4.60 (s, 2H), 7.33 (d, $J = 7.6$ Hz, 1H), 7.43 (t, $J = 7.6$ Hz, 1H), 7.49–7.59 (m, 4H), 7.80 (d, $J = 7.6$ Hz, 1H), 7.99 (d, $J = 7.6$ Hz, 1H), 8.36 (m, 2H). ^{13}C NMR (100.5 MHz) δ 41.4, 59.0, 123.5, 125.9, 126.7, 127.2, 127.5, 128.0, 128.2, 128.3, 128.5, 129.7, 130.5, 132.0, 137.5, 138.0, 139.9, 152.7, 192.4. IR (NaCl) ν 1615, 1633 cm^{-1} . HRMS (ESI-TOF) calcd for $\text{C}_{19}\text{H}_{16}\text{NO}$: 274.1226 $[\text{M} + \text{H}]^+$; found: 274.1225.

■ ASSOCIATED CONTENT

Supporting Information

^1H and ^{13}C NMR spectra of new compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

Support for this work by Grants CTQ2009-07175 and CTQ2012-31391 from the Ministerio de Economía y Competitividad, Spain, is gratefully acknowledged.

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